
REVIEWS

Circadian Regulation and Its Disorders in Parkinson's Disease Patients. Part 2: Experimental Models, alpha-Synuclein, and Melatonin

Yu. V. Ukraintseva^{a,*} and V. M. Kovalzon^{b,**}

^a*Institute of Higher Nervous Activity and Neurophysiology, Russian Academy of Sciences, Moscow, Russia*

^b*Severtsov Institute of Ecology and Evolution, Russian Academy of Sciences, Moscow, Russia*

*e-mail: ukraintseva@yandex.ru

**e-mail: kovalzon@sevin.ru

Received November 13, 2015

Abstract—Circadian disturbances related to Parkinson's disease are reviewed, and possible pathogenetic mechanisms are discussed. The role of dopaminergic system degeneration in the development of circadian dysfunction is emphasized. The accumulation of α -synuclein in the suprachiasmatic nucleus is considered as a possible mechanism of circadian dysfunction unrelated to dopamine deficiency. Data on the disbalance of dopamine and melatonin levels in Parkinson's disease patients and its role in disturbances of circadian rhythms of physiological processes are analyzed.

Keywords: Parkinson's disease, experimental models, circadian regulation, sleep-wakefulness, dopaminergic system, melatonin, α -synuclein

DOI: 10.1134/S0362119716050170

EXPERIMENTAL PARKINSON'S DISEASE (PD) MODELS DEMONSTRATE CIRCADIAN RHYTHM DISORDERS

Because of the large number of factors involved in the pathogenesis of PD (such as age, medical therapy, depression, anxiety, cognitive impairments, etc.), in clinical trials it is still impossible to distinguish those of them that affect the sleep–wakefulness cycle regulation at dopaminergic neurodegeneration. Similar studies were performed in animal models, in particular, using the proneurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine hydrochloride (MPTP) or the neurotoxin 6-oxydopamine, which lead to a selective destruction of the dopaminergic system.

Seven Californian drug addicts with severe symptoms of Parkinson's disease (discovered in 1983) were examined. On the basis of the results of these studies, the MPTP model was created. These young subjects practiced intravenous injections of crude meperidine, a synthetic analogue of heroin containing high concentrations of MPTP, a by-product of heroin synthesis. Thorough studies performed in subsequent years showed that MPTP, due to its high lipophilicity, easily passes through the barrier and penetrates into astrocytes, where it is converted into the 1-methyl-4-phenyl-pyridin ion (MPP⁺) by type B monoamine oxidase (MAO-B). This ion binds to dopamine transporter and thus gets into the mitochondria of

dopamine-containing neurons. In mitochondria, it inhibits complex-1 (bound to NADH ubiquinone oxidoreductase) of the mitochondrial electron transfer chain and thus uncouples oxidative phosphorylation. This, in turn, leads to disruption of adenosine triphosphate (ATP) production, increase in the extracellular calcium levels, and generation of free radicals/reactive oxygen species. The latter interact with cellular proteins, nucleic acids, lipids, and other molecules and cause cellular damage and, ultimately, the death of neurons (i.e., dopamine neurotoxicity symptoms) [1].

MPTP toxicity in different species of mammals varies widely and depends primarily on the level of MAO-B in the brain. This level is high in carnivorous and primates, low in rodents, and almost zero in rats (which determines their high resistance to disinfection procedures). For this reason, the relative doses of the toxin required to form the parkinsonian phenotype in cats and monkeys are 15 times lower than in mice. In rats, it is practically impossible to induce parkinsonism symptoms in chronic experiments with systemic MPTP administration [2].

Nevertheless, a model of Parkinson's disease, which is generally accepted today and which was validated by the International Society of Psychopharmacologists, was developed in black mice of the C57BL/6 strain. In this model, black C57BL/6 mice are systemically injected with the neurotoxin MPTP which

selectively destroys the dopaminergic system. The effect of this toxin depends on the dose and injection regimen. For this reason, the results of different studies of the effect of MPTP-induced destructions on the circadian rhythms in mice are contradictory. The authors of [3] detected no disturbances in circadian rhythms in mice after MPTP administration but observed severe destruction of dopaminergic neurons (approximately 50%) [3]. The authors of another study showed that MPTP affected the circadian rhythm of locomotor activity, lengthening the period of free locomotor activity (running in a wheel) compared to the control mice [4]. This was accompanied by a reduction in the number of dopaminergic neurons in the substantia nigra (SN) by 43%. It was also shown that MPTP-induced destructions caused changes in the architecture and efficiency of sleep in mice (in particular, increase or reduction in REM sleep) [5–9].

Simulation of dopaminergic system pathology is also widely used in other rodents and predators. For example, injections of the toxin oxydopamine-6 in rats lead to the loss of large amounts of dopaminergic neurons and disrupt both behavioral rhythms and clock gene expression rhythms [10]. In cats, MPTP injections reduce the REM sleep duration [11].

In the PD models in lower primates, caused by the destruction of the dopaminergic system with MPTP, an increased daytime sleepiness and sleep fragmentation were detected [12, 13]. In addition, MPTP was shown to significantly enhance the tonic muscle activity during REM sleep, which is indicative of a phenomenon similar to the REM sleep behavior disorder (RBD) [14]. Similarly to PD patients, dysregulation of REM sleep and increased daytime sleepiness occur before the motor symptoms [12, 14–16].

Increased daytime sleepiness and sleep fragmentation, possibly, indicate circadian dysfunction. Indeed, parkinsonism caused by MPTP administration in lower primates is associated not only with sleep disorders but also with changes in other circadian rhythms of behavior and physiological processes such as secretion of melatonin and prolactin [17] and rhythms of temperature and locomotor activity [18]. Hence, dopamine deficiency entails an overall circadian system disruption. A recent study [19] has shown that MPTP-induced destruction of the dopaminergic system leads to a decrease in amplitude, increase in fragmentation, and reduction in stability of the circadian rhythm of locomotor activity in animals kept in a light/dark regime. Under conditions of constant illumination, severe disturbances of rhythm and even its complete disorganization are observed, but the amplitude and phase of secretion of melatonin and cortisol remain unchanged. The authors concluded that after the dopaminergic system destruction the central “clock” in the suprachiasmatic nucleus (SCN) is retained. However, in the absence of stimulatory and inhibitory effects of light and dark the clock cannot

ensure the down-regulating effects on the clock genes of the striatum and the dopaminergic functions which control the locomotor behavior.

Another way to assess the role of the dopaminergic system in the sleep–wakefulness cycle regulation is the study of sleep and circadian rhythms in genetically modified animals lacking specific dopamine receptors or dopamine transporters. A decrease in the duration of wakefulness with a corresponding increase in the duration of both slow-wave and REM sleep as well as a sharp reduction in the δ -rhythm (0.75–2 Hz) power during slow-wave sleep was shown in D2 knockout mice [20]. In addition, an increase in the number of wakefulness episodes and a decrease in their length were observed, indicating the instability of wakefulness. The authors of this study concluded that D2 receptors play an important role in maintaining wakefulness.

Experiments in other models—in mice with deficient expression of the vesicular monoamine transporter (VMAT2) [21, 22] and in mice knockout for the dopamine transporter (DAT-KO mice) [23]—also showed that the disruption of the normal function of dopaminergic cells is associated with the appearance of non-motor symptoms, including sleep disorders and circadian rhythm disturbances.

Thus, different experimental PD models show that the dopaminergic system destruction and the dopamine deficiency lead to disturbances in the rhythms of clock gene expression, sleep–wakefulness, and voluntary locomotor activity.

PATHOGENIC MECHANISMS OF CIRCADIAN DISORDERS IN PD NOT RELATED TO DOPAMINE DEFICIENCY: ACCUMULATION OF α -SYNUCLEIN

Although the dopaminergic system destruction can lead to circadian dysfunction, the specific pathogenic mechanisms underlying circadian dysfunction in PD remain poorly understood.

Parkinson’s disease, in addition to the degeneration of the nigrostriatal dopaminergic system, can also be accompanied by the destruction of the neuroanatomical parts of the circadian system per se—afferent pathways to the SCN, SCN itself, and the descending peripheral efferents of the SCN. For example, the disturbance of the visual function accompanying PD which is due primarily to the degeneration of dopaminergic retinal networks and dopamine deficiency, is well known [24, 25]. As a result, the transmission of information about changes in illuminance is disturbed, which may affect the circadian rhythm maintenance in PD patients.

Data on the involvement of the hypothalamus in the pathological process in PD are ambiguous [26, 27], and the effect of PD on the structure and function of the SCN is still poorly understood. However, the

degeneration of this central pacemaker is considered as another possible mechanism explaining the circadian rhythm disturbances in PD patients [28]. Finally, the disturbances of efferent pathways in SCN may be responsible for the disruption of biological rhythms in PD (in particular, changes in the circadian rhythm of melatonin secretion in PD are known [29–32]).

The mechanisms that determine the circadian fluctuations in PD symptoms are not understood completely as well. These fluctuations may be determined in part by the fluctuations in the metabolism of dopamine, dopamine accumulation per night, or diurnal deactivation of receptors [33, 34].

Despite the fact that PD is primarily the result of degeneration of neurons in the SN which leads to subsequent reduction of the dopaminergic inputs to the striatum, a number of stem nuclei (locus coeruleus, raphe nuclei, dorsal motor vagal nucleus), cortical neurons (in particular, in the cingulate gyrus and entorhinal cortex), Meynert basal nucleus, and preganglionic sympathetic and parasympathetic neurons are also destroyed in this disease. The pathogenetic mechanisms of these diffuse disturbances in different parts of the brain are not understood completely; it was assumed that the accumulation of α -synuclein may play an important role in them [28].

Possible mechanisms of the effect of α -synuclein on circadian regulation can be considered using the mouse model of synucleinopathies as an example. One of the best studied models of PD and other synucleinopathies is the transgenic mouse strain *Thy1- α Syn* expressing human α -synuclein controlled by the *Thy-1* gene promoter [35]. Genetic mutations in the α -synuclein gene or duplication of this gene are closely associated with the PD family forms; polymorphism for this gene determines the risk of PD [36–40]. *Thy1- α Syn* transgenic mice exhibit a progressive disturbance of motor and non-motor functions similar to that observed in PD patients, including the impairment of olfaction and cognitive functions and disorders of the autonomic nervous system [41–43]. The study of circadian regulation in these mice showed [44] that they experience severe disorders of the circadian rhythm of locomotor activity: fragmentation, reduced rhythm amplitude due to the lower level of activity at night, and less clear beginning of the activity/rest periods (Fig. 1). Although the duration of the activity period, the circadian rhythm capture by the changes in the light regime, and the expression pattern of the clock gene *Per2* did not differ from the norm, the frequency of spontaneous action potentials in SCN neurons was significantly reduced during the daytime [28, 43].

It is known that the oscillatory clock mechanism of SCN is based on the rhythmic expression of the key clock genes, in particular, the *Per2* gene [44], which, in turn, controls the action potential fluctuations in SCN neurons projecting to other regions of the brain.

If the circadian clock in the SCN functions normally, the *Per2* expression level is increased in the daytime and decreased in the dark time of the day; as a result, the pulsation of SCN neurons will be at maximum in the daytime [45]. Since *Thy1- α Syn* mice retained a clear rhythm of fluctuations in the *Per2* gene expression in the SCN, it can be concluded that the disturbances in their periodicity were not the result of deficiency in molecular oscillations in the SCN or the pathology of its inputs, and the daytime decrease in the neuronal excitability in SCN is caused by some other factors.

A possible mechanism that mediates the decrease in the neuronal activity of SCN under conditions of α -synuclein overexpression may be the changes in the synaptic transmission. α -Synuclein is a presynaptic protein that regulates the release of synaptic vesicles, and its incorrect expression disrupts the synaptic transmission [46, 47]. Neurons in the nervous networks in the SCN release gamma-aminobutyric acid (GABA) as a neurotransmitter, and the majority of neurons receive a constant inflow of GABA signals [48–50]. Probably, α -synuclein overexpression affecting the synaptic transmission can shift the balance towards enhancement of inhibition in the SCN networks, which ultimately leads to the circadian symptoms in *Thy1- α Syn* mice.

An alternative explanation is based on the events that underlie the diurnal rhythms of spontaneous electrical activity of neurons in the SCN, whose depolarization increases in the daytime period. This relatively depolarized resting potential is the result of stimulatory effects implemented by multiple cationic currents [51, 52]. A decrease in the amplitude of these currents may reduce the daytime pulsation of SCN neurons [45, 53, 54]. It was also shown [55] that aging selectively disrupts potassium currents in the SCN which reduces the synchronicity of its cell population.

Although α -synuclein primarily plays a major role in the synapse in the processes of release and recycling of synaptic vesicles, there is evidence of its colocalization in the mitochondrial membrane [56, 57]. It was also shown that the mitochondrial function may be impaired due to incorrect α -synuclein expression [58] and, conversely, the mitochondrial proneurotoxin MPTP leads to α -synuclein accumulation [59].

On the other hand, the circadian system pathology may disrupt the function of mitochondria and induce oxidative stress. In particular, it was shown that a deletion in one of the key clock genes, *BMAL1*, leads to mitochondrial dysfunction, including an increase in the content of reactive oxygen species in peripheral organs [60–62]. Ample data have been accumulated showing that both the generation of reactive oxygen species and the production of cellular antioxidants are controlled by the circadian system [60, 63, 64] and that many parameters of the immune system have circadian oscillations (see, e.g., [65]).

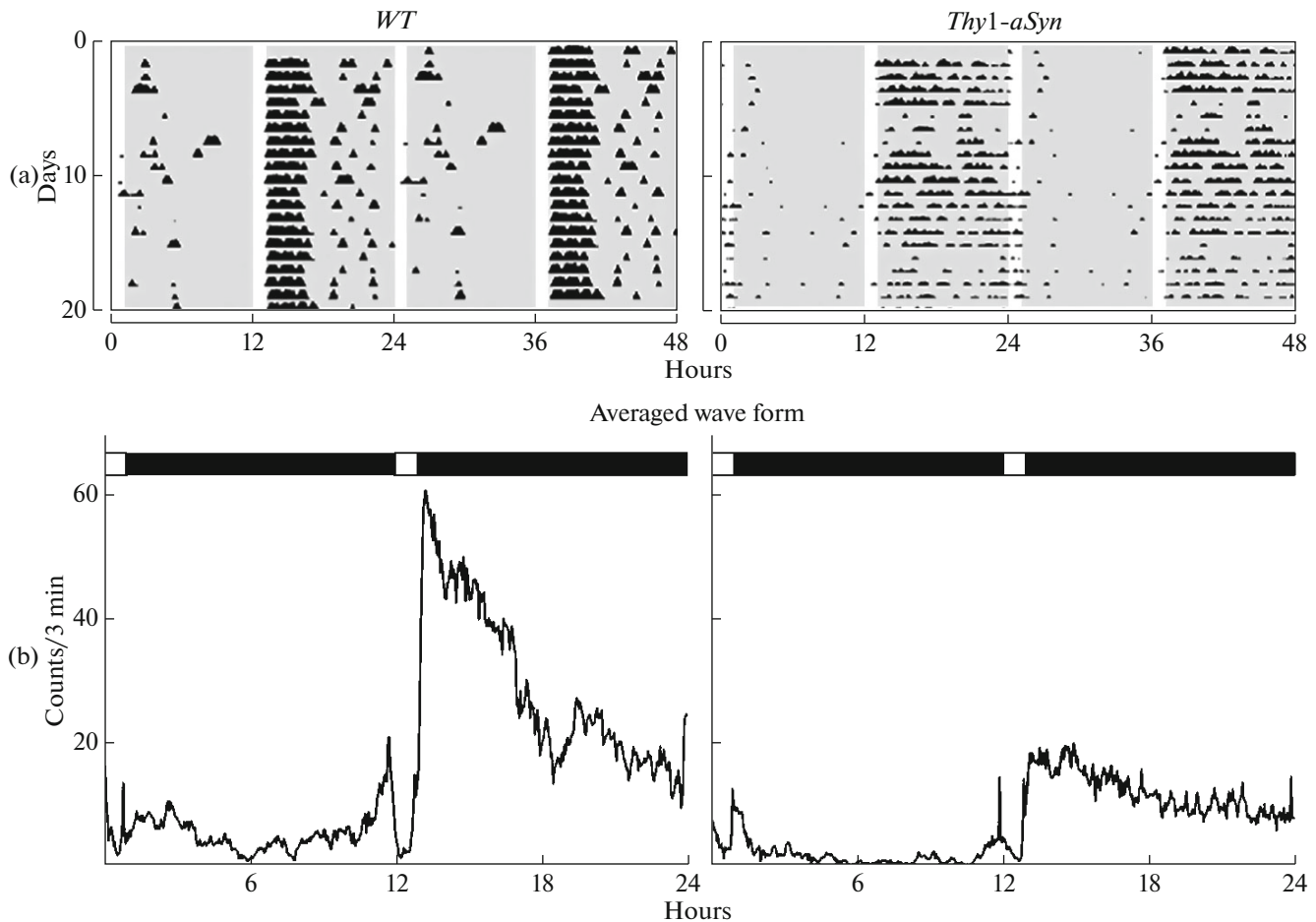


Fig. 1. Circadian rhythm disturbance may be the key component of the non-motor symptoms of Parkinson's disease. Data were obtained in the experimental model of transgenic mice overexpressing α -synuclein. (a) A representative plot of the wheel running intensity of the control (WT) and transgenic (*Thy1- α Syn*) mice. The abscissa axis shows hours, and the ordinate axis shows days. Animals that were kept under the 12 : 12 LD cycle, shifted to the fragmented photoperiodicity regimen 1 : 11 : 1 : 11 LD. Each horizontal row is a duplicated 24-h record of motor activity day by day (top to bottom). The shaded areas designate the dark periods of the day. (b) Averaged representative curves illustrating the dynamics of motor activity of control and mutant mice. The abscissa axis shows hours, and the ordinate axis shows the number of wheel turns per 3 min. In addition to the sharp decline in the level of activity, the transgenic mice showed disturbed timing of the onset of the periods of increased motor activity relative to switching off the light and an increased fragmentation of these periods. Thus, *Thy1- α Syn* transgenic mice overexpressing α -synuclein exhibit a smoothed circadian rhythm of voluntary motor activity [28, 43].

Thus, an enhancement of oxidative stress and inflammatory processes occurring as a result of circadian dysfunction in combination with α -synuclein aggregation can exacerbate the PD pathology. Therefore, the circadian system dysfunction can be considered as a risk factor for PD [28] (Fig. 2).

There are numerous data demonstrating that clear circadian rhythms are an essential component of a good health. Many studies have shown that disturbances in the circadian system cause a cluster of symptoms, including the cognitive deficit and memory problems [66, 67], metabolic disorders [68, 69], cardiovascular disorders [70, 71], gastrointestinal diseases [72, 73], and an increased risk of certain types of cancer [74]. Many of these symptoms were described in PD patients. It may be concluded that circadian dys-

function is not only a symptom of PD but also a deep (and, maybe, even a pathogenetic), component of the disease.

THE LEVEL OF MELATONIN AND DISTURBANCES IN CIRCADIAN REGULATION IN PD

The most important biological marker of the circadian system is melatonin, a hormone that is synthesized primarily in the pineal and plays an important role, in particular, in the regulation of sleep and seasonal biorhythms [75]. The synthesis of melatonin increases at the dark period of nycthemeron and is inhibited in the daytime.

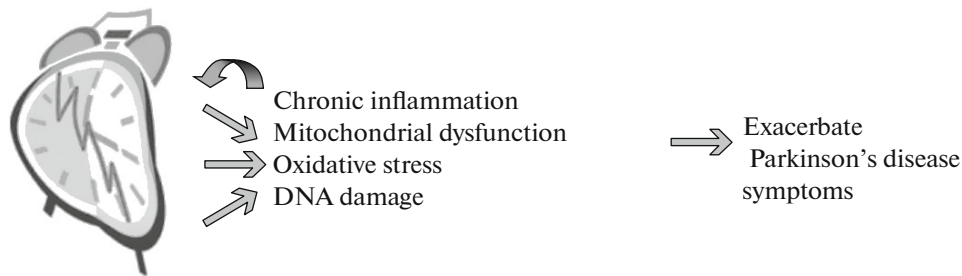


Fig. 2. Potential mechanism by which circadian dysfunction can exacerbate PD symptoms. The molecular clock of the body regulates the mitochondrial function, the generation and decay of reactive oxygen species, the DNA repair, and immune responses. Disturbances in the biological clock contribute to the development of chronic inflammatory processes, mitochondrial function disturbance, and DNA damage. All these processes are assumed to be involved in the development of Parkinson's disease symptoms and age-related changes in the brain. Circadian dysfunction caused by genetic factors or perturbations in the environment can accelerate the development of disorders in PD [28].

The study of the secretion of this hormone in PD showed, first of all, changes in the circadian rhythm of fluctuations in the melatonin concentration. In particular, the amplitude of the rhythm of melatonin secretion in PD patients decreases [29, 30]. The comparison of the treated (with levodopa and/or dopamine agonists) and untreated (newly diagnosed with PD) patients indicates a trend to phase advance in melatonin fluctuations in the treated patients [29, 31, 32]. In particular, according to [32], the usual bedtime and the time of the beginning of twilight melatonin secretion was not significantly different between the treated and untreated patients. However, the phase angle between the rhythms determined as the difference between the bedtime and the beginning of melatonin release was more than twice higher in the patients who received dopaminergic therapy compared to the untreated patients. According to the authors, the discordance of circadian regulation and sleep regulation in PD is caused by the use of dopaminergic drugs rather than the disease process itself.

The effect of PD on the overall level of secreted melatonin was also studied. Patients with the newly diagnosed PD were characterized by a decrease in the circulating melatonin level compared to the healthy age-matching subjects [76]. In another study, the patients who received dopaminergic drugs also had a lower melatonin concentration compared to the control group [30]. In addition, the effect of dopaminergic therapy on the melatonin level was revealed. In particular, the amount of circulating melatonin in the treated patients was higher compared to the untreated ones [32]. An increased daytime secretion of melatonin in the PD patients with the motor side effects of levodopa compared to the patients without the side effects and the untreated patients was detected [31]. In the study [77], PD patients treated with dopaminergic drugs had elevated levels of melatonin in the morning, and the melatonin level was positively correlated with the stage of the disease and did not depend on the dose of dopaminergic drugs. However, it should be noted

that the intensity of illumination, which is known to affect the level of melatonin, was controlled only in the single study [32].

The increased melatonin secretion in response to dopaminergic therapy can be explained by taking into account the data on the involvement of dopamine in the regulation of secretion of the pineal gland. In particular, the receptor D4 whose expression is dependent on the light intensity and obeys circadian fluctuations was identified in the pineal of rats [78]. Moreover, the secretion of serotonin and melatonin from the pineal gland is controlled by heteromerization of adrenergic and dopaminergic receptors, which also obeys the circadian rhythm. Using α_1 B-D4 and β_1 -D4 heteromeric receptors, dopamine inhibits the adrenergic receptor and blocks the synthesis of melatonin induced by adrenergic receptor ligands. This inhibition is not observed in the daytime, when D4 is not expressed. The identified heteromerization between the adrenergic and dopaminergic D4 receptors is a neurohumoral feedback mechanism by which the dopaminergic regulation of circadian inputs is realized [79].

In the study [32] despite the fact that the treated patients had a significantly longer time elapsed between the beginning of melatonin secretion and the bedtime, they showed no differences in the severity of insomnia and any other deviations in the sleep architecture. Some authors believe that melatonin exhibits somnogenic properties [80]. From this standpoint, the observed separation of these two events in time in the case of increased melatonin secretion seems paradoxical. Accordingly, the existence of certain forms of melatonin resistance in patients with PD was assumed which may supposedly explain the modest success in insomnia therapy in PD patients with the exogenous melatonin [81, 82]. For example, in one of the studies [81], no difference between the effect of 5 and 50 mg of melatonin on the sleep quality and daytime sleepiness in such patients was detected (it should be noted that both doses used were too high to demonstrate the potential somnogenic effect [75]). Data on the reduc-

tion in expression of melatonin receptor types 1 and 2 in the striatum and other brain regions that are affected in PD indirectly indicate the development of such resistance [83].

However, to our opinion this is not the main cause. Melatonin is not a “sleep hormone,” because it is released at night in diurnal, nocturnal, as well as crepuscular mammals. Its formation requires two factors: (a) the absence of bright lighting and (b) the absence of activity of SCN neurons. In the diurnal (daytime) mammals, including humans, the release of melatonin by the pineal gland, indeed, coincides with the usual sleep hours, which makes attractive the hypothesis of a causal relationship between these two phenomena. It was shown that an increase in the daytime level of systemic melatonin, which was caused by intravenous infusion of tryptophan (melatonin precursor) or 5-methoxypsoralen, which suppressed its degradation, enhanced subjective sleepiness and shortened the latent period of sleep in healthy subjects. Conversely, the suppression of melatonin production by administration of β -blockers destroyed the sleep architecture. In three species of diurnal monkeys (*Macaca mulatta*, *M. nemestrina*, and *M. fascicularis*), evening oral administration of low (“physiological”) doses of melatonin decreased the latency and lengthened the night rest period, which can be regarded as a mild somnogenic effect [75].

In humans, the elevation of melatonin levels is not a mandatory signal to falling asleep. In the majority of healthy subjects, the administration of melatonin at “physiological” doses causes only a mild sedative effect enabling the general relaxation and reducing the responsiveness to normal surrounding stimuli which leads to calm wakefulness and gradual falling asleep. Unlike the strong “night” sedatives and hypnotics of the benzodiazepine series melatonin does not cause the sensation of an unbearable fatigue and irresistible thrust to sleep. If this subject is motivated he can easily overcome the “somnogenic” properties of melatonin. Both objective (assessed on the basis of polysomnograms) and subjective (based on the reports of healthy subjects and patients with insomnia) pharmacological characteristics of the “classical” benzodiazepine sedatives and hypnotics, on the one hand, and melatonin, on the other hand, drastically differ [75].

On the basis of the correlation between the subjectively sensed and objectively confirmed increase in evening sleepiness, on the one hand, and the beginning of the increase in the melatonin level in blood, on the other hand, it was assumed that, in humans, melatonin creates the “predisposition to sleep” and inhibits wakefulness mechanisms rather than directly affects the somnogenic structures. Due to the high saturation of the SCN and the adjacent areas of the preoptic area with the high-affinity melatonin receptors, this hormone, along with a number of other physical (bright light) and biochemical (the neu-

rotransmitters glutamic acid and serotonin as well as the neuropeptides neuropeptide tyrosine (NPY) and substance P (SP) factors, can exert a strong modulating effect on the activity of the main oscillator in mammals including humans. For example, when administered in the morning, melatonin causes a delay in the circadian phase in humans; however, when administered in the evening, it, conversely, causes advanced phase shift. These phase shifts in humans do not exceed 30–60 min per day. Thus, a daily administration of melatonin can help to achieve a forward or backward shift in the activity–rest circadian cycle in humans for several hours [75].

In general, the data on the therapeutic effect of exogenous melatonin regarding the correction of sleep disorders are ambiguous. A number of studies have shown that melatonin decreases the problems with sleep initiation and the nocturnal activity in the elderly [84–86]. The therapeutic effects of exogenous melatonin in RBD were also repeatedly observed [87–89]. However, the hypnotic effect of melatonin was not always confirmed [90]. Furthermore, this effect proved to be limited: it was shown that melatonin improved the subjective estimation of sleep in patients, but the objective improvement in sleep quality was minimal [81, 82]. However, it was assumed that these failures may have been related to the short half-life of melatonin when it is used in inappropriately high doses [91].

The studies with an oral administration of “physiological” (0.1–0.3 mg) doses of melatonin during the day increasing the plasma melatonin level to 50–120 pg/mL which roughly corresponds to its nighttime level in adult healthy subjects showed only a very small, though significant somnogenic effect. This effect was manifested in strengthening the subjective sleepiness and shortening the latent period of the first and second stages. Evening melatonin administration improved the nocturnal sleep characteristics and shortened latency in patients with insomnia but almost did not alter the sleep structure in healthy subjects [75]. There is evidence that the effect of melatonin is bell-shaped, similarly to the effects of serotonin and dopamine, which at high concentrations cause the “paradoxical” effects.

There is also evidence that melatonin increases motor symptoms in PD patients [92–94] and may exacerbate nightmares and moving activity during sleep [92]. Since the relationship between the elevated levels of melatonin and more severe parkinsonism symptoms in patients [29, 77, 95] and experimental animals were noted, adverse effects of melatonin on motor function were revealed [96, 97]. In view of above, further studies of the effect of exogenous melatonin in PD patients are required.

Interestingly, an epidemiological study based on data from 84794 nurses showed that shift work with night duties reduces (!) the risk of PD development by

50% [98]. Since it was shown that the concentration of circulating melatonin at shift work decreases [99], the lower risk of PD can be, probably, explained by the decrease in the melatonin level due to the prolonged effect of light during night duties.

Light, the main synchronizing factor for the human circadian system, is increasingly widely used to correct various somnological and neuropsychiatric disorders, including the circadian rhythm disorders, seasonal affective disorders, and dementia [100]. A therapeutic effect of light therapy was also found in PD [94, 101–103]. Dopamine is the main neurotransmitter that mediates the input of signals about illumination changes to the retinal circadian clock which sends direct projections to the SCN. Light stimulates the synthesis, turnover, and release of dopamine in the retina [104, 105], so that an exposure to bright light, apparently, makes it possible to compensate for the dopamine deficiency. In addition to the positive effect of bright light on the mood, it was shown to reduce the severity of bradykinesia, rigidity, and dyskinesias [101–103]. The well-known therapeutic effects of REM sleep deprivation on the motor symptoms of PD [106–108] can also be determined by the activating effect of light rather than the deprivation itself [109].

It is known that melatonin and dopamine in the retina are in a reciprocal relationship and play opposite regulatory roles in its adaptation to the day/night conditions. The synthesis of melatonin is suppressed by light, and in the absence of normal amount of light in the environment, the synthesis and secretion of melatonin increase, whereas the level of dopamine decreases [110, 111]. A similar balance between dopamine and melatonin exists in the pineal gland: the concentration of dopamine is increased in daytime, whereas the concentration of melatonin is increased at night [112].

The relationship between the visual impairment and dopaminergic degeneration is also well-known. In particular, disturbances in the visual function in PD such as elongation of evoked potential latencies, reduction in contrast sensitivity, and electroretinogram pattern changes are associated with the retinal dopamine deficiency [113, 114] and correlate with the severity of clinical manifestations of PD [115]. Possibly, the visual deficiency, which develops in PD, is accompanied by a decrease in the effect of light on the retina, which, in turn, leads to an increased daytime melatonin secretion and further disturbs the balance between dopamine and melatonin. Moreover, age-related changes in the visual function due to cataract, macular degeneration, and ganglion cell death can shift this balance, thereby contributing to the development of PD.

There is a standpoint according to which various functional disorders in PD are caused primarily by the imbalance between dopamine and melatonin rather than by the dopamine deficiency [109]. According to

Fig. 3, melatonin and dopamine in the retina and pineal gland are in reciprocal relationships. During the day, the melatonin level decreases whereas the dopamine level increases (a). Conversely, at night the melatonin level increases whereas the dopamine level decreases (b). However, normally these two systems are functionally coupled (connected by the thin black line in Fig. 3), and their changes in natural conditions are balanced. In PD, the relationship between dopamine and melatonin is disturbed (c), and the dopamine level decreases (at severe symptoms, to 20% of the norm), whereas the melatonin level increases. Long-term dopamine replacement therapy returns the ratio between the levels of dopamine and melatonin to nearly normal values (d). However, long-term dopamine replacement therapy causes an internal compensatory response aimed at restoring the dopamine–melatonin balance which gradually raises the dopamine level above the melatonin level and nullifies the result of treatment (e). A further increase in the melatonin level caused by increasing doses of dopamine-containing drugs throws the entire system into chaos, and the dopamine level continues to increase against the background of the increase in the melatonin level, once again imbalancing the system and causing dyskinesia (f). It was shown that the administration of exogenous melatonin improves the involuntary movements and reduces the severity of dyskinesia induced by an overdose of dopaminergic drugs (g). This is probably due to the return of dopamine and melatonin to the reciprocal functional relationship but at a higher level than in the healthy subjects. This assumption is confirmed by the reports that, although melatonin alleviates dyskinesia it does not improve the symptoms of the disease in general. A functional balance between dopamine and melatonin can be reached most easily using melatonin antagonists (epiphysectomy, bright light, or melatonin receptor blockade) against the background of a slight or moderate increase in the dopamine level (h). The maintenance of this kind of balance between dopamine and melatonin seems to be even more important than the dopamine replacement itself [109].

Dopamine replacement therapy also does not always allow restoring the delicate dynamic balance between the content of dopamine and melatonin, as evidenced by the “on–off” phenomena, which appear in the patients receiving long-term therapy with dopaminergic drugs and are accompanied by an elevated level of melatonin [29]. Given the fact that melatonin can inhibit the release of dopamine in various brain areas (such as the ventral hippocampus, pons, medulla, hypothalamus, and striatum [116]), an imbalance between dopamine and melatonin may exacerbate the disease.

From this standpoint, the therapeutic effect of bright light on PD patients may be due to the restoration of circadian rhythmicity and normalization of the balance between melatonin and dopamine [109,

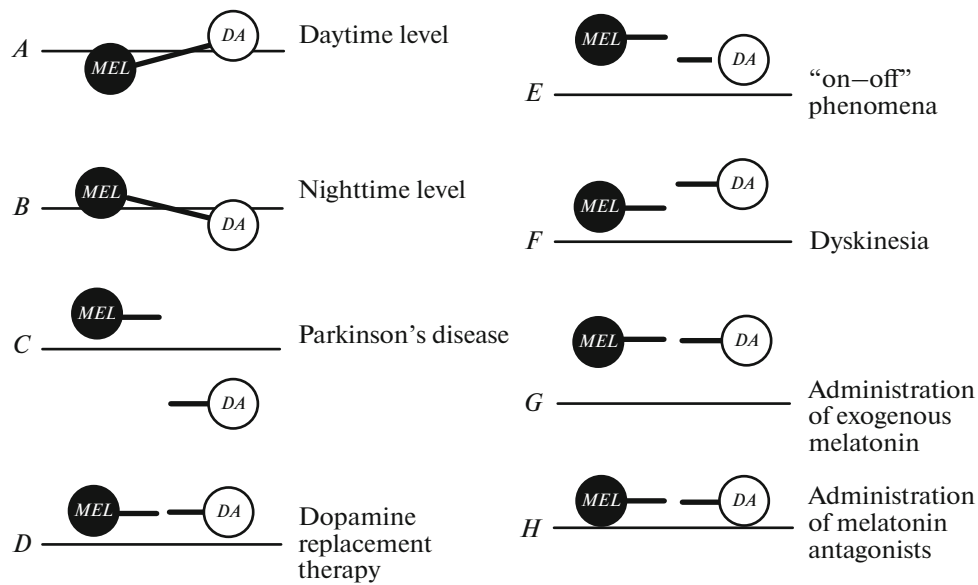


Fig. 3. Balance of dopamine (DA) and melatonin (MEL) in normal state and in PD [109] (for explanation, see the text).

117]. This assumption was confirmed by the study of the effect of melatonin on experimental PD symptoms, which showed that a slow injection of melatonin into the brain ventricles enhances the symptoms of parkinsonism, whereas the removal of the epiphysis or keeping animals under constant light conditions caused remission [96]. Thus, a decrease in the bioavailability of endogenous melatonin may alleviate the symptoms of PD. For this reason, melatonin (which exhibited neuroprotective properties in many studies [118]) should be used with caution in PD.

Thus, PD is accompanied by the circadian dysfunction, which significantly impairs the quality of life of patients. Its possible causes include dopamine deficiency, α -synuclein aggregation, imbalance between melatonin and dopamine secretion, as well as the neurodegenerative process in the SCN and its afferents. Studies have shown that the disruption of circadian rhythms observed in PD patients is pathogenetically associated with PD. On the other hand, the disturbance of circadian regulation itself adversely affects all functions of the body and may exacerbate the neurodegenerative processes.

ACKNOWLEDGMENTS

This study was supported by the Russian Foundation for Basic Research, project no. 16-04-01403a.

REFERENCES

1. Yokoyama, H., Kuroiwa, H., Kasahara, J., and Araki, T., Neuropharmacological approach against MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine)-induced mouse model of Parkinson's disease, *Acta Neurobiol. Exp.*, 2011, vol. 71, no. 2, p. 269.
2. Giovanni, A., Sieber, B.A., Heikkila, R.E., and Sonsalla, P.K., Studies on species sensitivity to the dopaminergic neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine. Part 1: Systemic administration, *J. Pharmacol. Exp. Ther.*, 1994, vol. 270, no. 3, p. 1000.
3. Laloux, C., Derambure, P., Kreisler, A., et al., MPTP-treated mice: long-lasting loss of nigral TH-ir neurons but not paradoxical sleep alterations, *Exp. Brain Res.*, 2008, vol. 186, no. 4, p. 635.
4. Tanaka, M., Yamaguchi, E., Takahashi, M., et al., Effects of age-related dopaminergic neuron loss in the substantia nigra on the circadian rhythms of locomotor activity in mice, *Neurosci. Res.*, 2012, vol. 74, nos. 3–4, p. 210.
5. Kovalzon, V.M., Ugrumov, M.V., Pronina, T.S., et al., Early stages of Parkinson's disease: Comparative characteristics of sleep–wakefulness cycle in patients and model animals, *Hum. Physiol.*, 2015, vol. 41, no. 6, p. 667.
6. Laloux, C., Derambure, P., Houdayer, E., et al., Effect of dopaminergic substances on sleep/wakefulness in saline- and MPTP-treated mice, *J. Sleep Res.*, 2008, vol. 17, p. 101.
7. Lima, M.M.S., Andersen, M.L., Reksidler, A.B., et al., The role of the substantia nigra pars compacta in regulating sleep patterns in rats, *PLoS One*, 2007, vol. 2, no. 6, p. e513.
8. Monaca, C., Laloux, C., Jacquesson, J.M., et al., Vigilance states in a Parkinsonian model, the MPTP mouse, *Eur. J. Neurosci.*, 2004, vol. 20, p. 2474.
9. Manolov, A.I., Dolgikh, V.V., Ukraintseva, Yu.V., et al., Changes in motor activity and the sleep–waking cycle in an MPTP model of Parkinson's disease in mice, *Hum. Physiol.*, 2014, vol. 46, no. 4, p. 467.
10. Gravotta, L., Gavrila, A.M., Hood, S., and Amir, S., Global depletion of dopamine using intracerebroventricular 6-hydroxydopamine injection disrupts normal

- circadian wheel-running patterns and PERIOD2 expression in the rat forebrain, *J. Mol. Neurosci.*, 2011, vol. 45, no. 2, p. 162.
11. Pungor, K., Papp, M., Kekesi, K., and Juhasz, G., A novel effect of MPTP: the selective suppression of paradoxical sleep in cats, *Brain Res.*, 1990, vol. 525, p. 310.
 12. Barraud, Q., Lambrecq, V., Forni, C., et al., Sleep disorders in Parkinson's disease: the contribution of the MPTP non-human primate model, *Exp. Neurol.*, 2009, vol. 219, no. 2, p. 574.
 13. Hyacinthe, C., Barraud, Q., Tison, F., et al., D1 receptor agonist improves sleep-wake parameters in experimental parkinsonism, *Neurobiol. Dis.*, 2014, vol. 63, p. 20.
 14. Verhave, P.S., Jongasma, M.J., Berg, R.M., et al., REM sleep behavior disorder in the marmoset MPTP model of early Parkinson disease, *Sleep*, 2011, vol. 34, no. 8, p. 1119.
 15. Vezoli, J., Fifel, K., Leviel, V., et al., Early presymptomatic and long-term changes of rest activity cycles and cognitive behavior in a MPTP-monkey model of Parkinson's disease, *PLoS One*, 2011, vol. 6, no. 8, e23952.
 16. Almirall, H., Pigarev, I., de la Calzada, M.D., et al., Nocturnal sleep structure and temperature slope in MPTP treated monkeys, *J. Neural Transm.*, 1999, vol. 106, nos. 11–12, p. 1125.
 17. Barcia, C., Bautista, V., Sanchez-Bahillo, A., et al., Circadian determinations of cortisol, prolactin and melatonin in chronic methyl-phenyl-tetrahydropyridine treated monkeys, *Neuroendocrinology*, 2003, vol. 78, no. 2, p. 118.
 18. Almirall, H., Bautista, V., Sanchez-Bahillo, A., and Trinidad-Herrero, M., Ultradian and circadian body temperature and activity rhythms in chronic MPTP treated monkeys, *Neurophysiol. Clin.*, 2001, vol. 31, no. 3, p. 161.
 19. Fifel, K., Vezoli, J., Dzahini, K., et al., Alteration of daily and circadian rhythms following dopamine depletion in MPTP treated non-human primates, *PLoS One*, 2014, vol. 9, no. 1, e86240.
 20. Qu, W.M., Xu, X.H., Yan, M.M., et al., Essential role of dopamine D2 receptor in the maintenance of wakefulness, but not in homeostatic regulation of sleep, in mice, *J. Neurosci.*, 2010, vol. 30, no. 12, p. 4382.
 21. Taylor, T.N., Caudle, W.M., Shepherd, K.R., et al., Nonmotor symptoms of Parkinson's disease revealed in an animal model with reduced monoamine storage capacity, *J. Neurosci.*, 2009, vol. 29, no. 25, p. 8103.
 22. Taylor, T.N., Caudle, W.M., and Miller, G.W., VMAT2-deficient mice display nigral and extranigral pathology and motor and nonmotor symptoms of Parkinson's disease, *Parkinson's Dis.*, 2011, artic. 124165.
 23. Dzirasa, K., Ribeiro, S., Costa, R., et al., Dopaminergic control of sleep-wake states, *J. Neurosci.*, 2006, vol. 26, no. 41, p. 10577.
 24. Harnois, C. and Di Paolo, T., Decreased dopamine in the retinas of patients with Parkinson's disease, *Invest. Ophthalmol. Visual Sci.*, 1990, vol. 31, no. 11, p. 2473.
 25. Archibald, N.K., Clarke, M.P., Mosimann, U.P., and Burn, D.J., The retina in Parkinson's disease, *Brain*, 2009, vol. 132, p. 1128.
 26. Matzuk, M.M. and Saper, C.B., Preservation of hypothalamic dopaminergic neurons in Parkinson's disease, *Ann. Neurol.*, 1985, vol. 18, no. 5, p. 552.
 27. Politis, M., Piccini, P., Pavese, N., et al., Evidence of dopamine dysfunction in the hypothalamus of patients with Parkinson's disease: an in vivo 11C-raclopride PET study, *Exp. Neurol.*, 2008, vol. 214, no. 1, p. 112.
 28. Willison, L.D., Kudo, T., Loh, D.H., et al., Circadian dysfunction may be a key component of the non-motor symptoms of Parkinson's disease: Insights from a transgenic mouse model, *Exp. Neurol.*, 2013, vol. 243, p. 57.
 29. Bordet, R., Devos, D., Brique, S., et al., Study of circadian melatonin secretion pattern at different stages of Parkinson's disease, *Clin. Neuropharmacol.*, 2003, vol. 26, no. 2, p. 65.
 30. Videnovic, A., Noble, C., Reid, K.J., et al., Circadian melatonin rhythm and excessive daytime sleepiness in Parkinson disease, *JAMA Neurol.*, 2014, vol. 71, no. 4, p. 463.
 31. Fertl, E., Auff, E., Doppelbauer, A., and Waldhauser, F., Circadian secretion pattern of melatonin in de novo Parkinsonian patients: Evidence for phase-shifting properties of L-dopa, *J. Neural Transm.: Parkinson's Dis. Dementia Sect.*, 1993, vol. 5, no. 3, p. 227.
 32. Bolitho, S.J., Naismith, S.L., Rajaratnam, S.M., et al., Disturbances in melatonin secretion and circadian sleep-wake regulation in Parkinson disease, *Sleep Med.*, 2014, vol. 15, no. 3, p. 342.
 33. Khaldy, H., Leon, J., Escames, G., et al., Circadian rhythms of dopamine and dihydroxyphenyl acetic acid in the mouse striatum: Effects of pinealectomy and of melatonin treatment, *Neuroendocrinology*, 2002, vol. 75, no. 3, p. 201.
 34. Weber, M., Lauterburg, T., Tobler, I., and Burgunder, J.M., Circadian patterns of neurotransmitter related gene expression in motor regions of the rat brain, *Neurosci. Lett.*, 2004, vol. 358, p. 17.
 35. Rockenstein, E., Mallory, M., Hashimoto, M., et al., Differential neuropathological alterations in transgenic mice expressing alpha-synuclein from the platelet-derived growth factor and Thy-1 promoters, *J. Neurosci. Res.*, 2002, vol. 68, p. 568.
 36. Cookson, M.R., Alpha-synuclein and neuronal cell death, *Mol. Neurodegener.*, 2009, vol. 4, p. 9.
 37. Pankratz, N., Wilk, J.B., Latourelle, J.C., et al., Genome-wide association study for susceptibility genes contributing to familial Parkinson disease, *Hum. Genet.*, 2009, vol. 124, p. 593.
 38. Ritz, B., Rhodes, S.L., Bordelon, Y., and Bronstein, J., α -Synuclein genetic variants predict faster motor symptom progression in idiopathic Parkinson disease, *PLoS One*, 2012, vol. 7, no. 5, e36199.
 39. Simón-Sánchez, J., Schulte, C., Bras, J.M., et al., Genome wide association study reveals genetic risk underlying Parkinson's disease, *Nat. Genet.*, 2009, vol. 41, no. 12, p. 1308.
 40. Vekrellis, K., Xilouri, M., Emmanouilidou, E., et al., Pathological roles of α -synuclein in neurological disorders, *Lancet Neurol.*, 2011, vol. 10, no. 11, p. 1015.

41. Fleming, S.M., Salcedo, J., Hutson, C.B., et al., Behavioral effects of dopaminergic agonists in transgenic mice overexpressing human wildtype alpha-synuclein, *Neuroscience*, 2006, vol. 142, p. 1245.
42. Fleming, S.M. and Chesselet, M.F., *Modeling Non-Motor Symptoms of Parkinson's Disease in Genetic Mouse Models*, Groenewegen, H.J., Voorn, P., Berendse, H.W., et al., Eds., vol. IX: *Basal Ganglia*, New York: Springer, 2009, p. 483.
43. Kudo, T., Loh, D.H., Truong, D., et al., Circadian dysfunction in mouse model of Parkinson's disease, *Exp. Neurol.*, 2011, vol. 232, no. 1, p. 66.
44. Hastings, M., Reddy, A., and Maywood, E., A clockwork web: circadian timing in brain and periphery, in health and disease, *Nat. Rev. Neurosci.*, 2003, vol. 4, p. 649.
45. Colwell, C.S., Linking neural activity and molecular oscillations in the SCN, *Nat. Rev. Neurosci.*, 2011, vol. 12, p. 553.
46. Burrè, J., Sharma, M., Tsetsenis, T., et al., Alpha-synuclein promotes SNARE-complex assembly in vivo and in vitro, *Science*, 2010, vol. 329, no. 5999, p. 1663.
47. Cabin, D.E., Shimazu, K., Murphy, D., et al., Synaptic vesicle depletion correlates with attenuated synaptic responses to prolonged repetitive stimulation in mice lacking alpha-synuclein, *J. Neurosci.*, 2002, vol. 22, no. 20, p. 8797.
48. Jiang, Z.G., Yang, Y., Liu, Z.P., and Allen, C.N., Membrane properties and synaptic inputs of suprachiasmatic nucleus neurons in rat brain slices, *J. Physiol.*, 1997, vol. 499, p. 141.
49. Strecker, G.J., Wuarin, J.P., and Dudek, F.E., GABA-mediated local synaptic pathways connect neurons in the rat suprachiasmatic nucleus, *J. Neurophysiol.*, 1997, vol. 78, p. 2217.
50. Itri, J., Michel, S., Waschek, J.A., and Colwell, C.S., Circadian rhythm in inhibitory synaptic transmission in the mouse suprachiasmatic nucleus, *J. Neurophysiol.*, 2004, vol. 92, p. 311.
51. Jackson, A.C., Yao, G.L., and Bean, B.P., Mechanism of spontaneous firing in dorsomedial suprachiasmatic nucleus neurons, *J. Neurosci.*, 2004, vol. 24, p. 7985.
52. Kononenko, N.I., Medina, I., and Dudek, F.E., Persistent subthreshold voltage dependent cation single channels in suprachiasmatic nucleus neurons, *Neuroscience*, 2004, vol. 129, no. 1, p. 85.
53. Kudo, T., Loh, D.H., Kuljis, D., et al., Fast delayed rectifier potassium current: critical for input and output of the circadian system, *J. Neurosci.*, 2011, vol. 31, no. 8, p. 2746.
54. Montgomery, J.R. and Meredith, A.L., Genetic activation of BK currents in vivo generates bidirectional effects on neuronal excitability, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, vol. 109, no. 46, p. 18997.
55. Farajnia, S., Michel, S., Deboer, T., et al., Evidence for neuronal desynchrony in the aged suprachiasmatic nucleus clock, *J. Neurosci.*, 2012, vol. 32, no. 17, p. 5891.
56. Li, W.W., Yang, R., Guo, J.C., et al., Localization of alpha-synuclein to mitochondria within midbrain of mice, *NeuroReport*, 2007, vol. 18, no. 15, p. 1543.
57. Nakamura, K., Nemani, V.M., Wallender, E.K., et al., Optical reporters for the conformation of alpha-synuclein reveal a specific interaction with mitochondria, *J. Neurosci.*, 2008, vol. 28, no. 47, p. 12305.
58. Martin, L.J., Pan, Y., Price, A.C., et al., Parkinson's disease alpha-synuclein transgenic mice develop neuronal mitochondrial degeneration and cell death, *J. Neurosci.*, 2006, vol. 26, no. 1, p. 41.
59. Purisai, M.G., McCormack, A.L., Langston, W.J., et al., Alpha-synuclein expression in the substantia nigra of MPTP-lesioned non-human primates, *Neurobiol. Dis.*, 2005, vol. 20, no. 3, p. 898.
60. Khapre, R.V., Kondratova, A.A., Susova, O., and Kondratov, R.V., Circadian clock protein BMAL1 regulates cellular senescence in vivo, *Cell Cycle*, 2011, vol. 10, no. 23, p. 4162.
61. Kondratov, R.V., Kondratova, A.A., Gorbacheva, V.Y., et al., Early aging and age-related pathologies in mice deficient in BMAL1, the core component of the circadian clock, *Genes Dev.*, 2006, vol. 20, no. 14, p. 1868.
62. Lee, J., Kim, M.S., Li, R., et al., Loss of Bmal1 leads to uncoupling and impaired glucose-stimulated insulin secretion in β -cells, *Islets*, 2011, vol. 3, no. 6, p. 381.
63. Antoch, M.P. and Kondratov, R.V., Circadian proteins and genotoxic stress response, *Circ. Res.*, 2010, vol. 106, p. 68.
64. Kondratova, A.A. and Kondratov, R.V., The circadian clock and pathology of the ageing brain, *Nat. Rev. Neurosci.*, 2012, vol. 13, no. 5, p. 325.
65. Logan, R.W. and Sarkar, D.K., Circadian nature of immune function, *Mol. Cell. Endocrinol.*, 2012, vol. 349, no. 1, p. 82.
66. Gerstner, J.R. and Yin, J.C., Circadian rhythms and memory formation, *Nat. Rev. Neurosci.*, 2010, vol. 11, no. 8, p. 577.
67. Loh, D., Navarro, J., Hagopian, A., et al., Rapid changes in the light/dark cycle disrupt memory of conditioned fear in mice, *PLoS One*, 2010, vol. 5, e12546.
68. Gale, J.E., Cox, H.I., Qian, J., et al., Disruption of circadian rhythms accelerates development of diabetes through pancreatic beta-cell loss and dysfunction, *J. Biol. Rhythms*, 2011, vol. 26, no. 5, p. 423.
69. Marcheva, B., Ramsey, K., Buhr, E., et al., Disruption of the clock components CLOCK and BMAL1 leads to hypo insulinaemia and diabetes, *Nature*, 2010, vol. 466, no. 7306, p. 627.
70. Bray, M., Shaw, C., Moore, M., et al., Disruption of the circadian clock within the cardiomyocyte influences myocardial contractile function, metabolism, and gene expression, *Am. J. Physiol. Heart Circ. Physiol.*, 2008, vol. 294, no. 2, p. 1036.
71. Scheer, F.A., Hilton, M.F., Mantzoros, C.S., and Shea, S.A., Adverse metabolic and cardiovascular consequences of circadian misalignment, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, vol. 106, no. 11, p. 4453.
72. Jain, S., Multi-organ autonomic dysfunction in Parkinson disease, *Parkinsonism Relat. Disord.*, 2011, vol. 17, no. 2, p. 77.
73. Gimble, J.M. and Floyd, Z.E., Metabolism: what causes the gut's circadian instincts?, *Curr. Biol.*, 2011, vol. 21, no. 16, p. 24.

74. Monsees, G.M., Kraft, P., Hankinson, S.E., et al., Circadian genes and breast cancer susceptibility in rotating shiftworkers, *Int. J. Cancer*, 2012, vol. 131, no. 11, p. 2547.
75. Koval'zon, V.M., *Osnovy somnologii. Fiziologiya i neirokimiya tsikla boдрstvovanie-son (Foundations of Somnology: Physiology and Neurochemistry of the Wake-Sleep Cycle)*, Moscow: Binom—Laboratoriya Znaniy, 2011.
76. Breen, D.P., Vuono, R., Nawarathna, U., et al., Sleep and circadian rhythm in early Parkinson disease, *JAMA Neurol.*, 2014, vol. 71, no. 5, p. 589.
77. Lin, L., Du, Y., Yuan, S., et al., Serum melatonin is an alternative index of Parkinson's disease severity, *Brain Res.*, 2014, vol. 1547, p. 43.
78. Kim, J.-S., Bailey, M.J., Weller, J.L., et al., Thyroid hormone and adrenergic signaling interact to control pineal expression of the dopamine receptor D4 gene (*Drd4*), *Mol. Cell. Endocrinol.*, 2010, vol. 314, no. 1, p. 128.
79. González, S., Moreno-Delgado, D., Moreno, E., et al., Circadian-related heterodimerization of adrenergic and dopamine D₄ receptors modulates melatonin synthesis and release in the pineal gland, *PLoS Biol.*, 2012, vol. 10, no. 6, e1001347.
80. Jan, J.E., Reiter, R.J., Wong, P.K., et al., Melatonin has membrane receptor-independent hypnotic action on neurons: a hypothesis, *J. Pineal Res.*, 2011, vol. 50, no. 3, p. 233.
81. Dowling, G.A., Mastick, J., Colling, E., et al., Melatonin for sleep disturbances in Parkinson's disease, *Sleep Med.*, 2005, vol. 6, no. 5, p. 459.
82. Medeiros, C.A., Carvalho de Bruin, P.F., Lopes, L.A., et al., Effect of exogenous melatonin on sleep and motor dysfunction in Parkinson's disease. A randomized, double blind, placebo-controlled study, *J. Neurol.*, 2007, vol. 254, no. 4, p. 459.
83. Adi, N., Mash, D.C., Ali, Y., et al., Melatonin MT1 and MT2 receptor expression in Parkinson's disease, *Med. Sci. Monit.*, 2010, vol. 16, no. 2, p. BR61. <http://www.medscimonit.com/fulltxt.php?ICID=878353>.
84. Garfinkel, D., Laudon, M., and Zisapel, N., Improvement of sleep quality by controlled release melatonin in benzodiazepine-treated elderly insomniacs, *Arch. Gerontol. Geriatr.*, 1997, vol. 24, no. 2, p. 223.
85. MacFarlane, J.C., Cleghorn, J.M., Brown, G.M., and Streiner, D.L., The effects of exogenous melatonin on the total sleep time and daytime alertness of chronic insomniacs: a preliminary study, *Biol. Psychiatry*, 1991, vol. 30, no. 4, p. 371.
86. Nodel', M.R., Current possibilities of therapy for sleep disorders in Parkinson's disease, *Nevrol. Neiropsikhiatr. Psikhosomatika*, 2013, no. 2, p. 30.
87. Kunz, D. and Mahlberg, R., A two-part, double-blind, placebo-controlled trial of exogenous melatonin in REM sleep behavior disorder, *J. Sleep Res.*, 2010, vol. 19, no. 4, p. 591.
88. Kunz, D. and Bes, F., Melatonin as a therapy in REM sleep behavior disorder patients: an open-labeled pilot study on the possible influence of melatonin on REM-sleep regulation, *Mov. Disord.*, 1999, vol. 14, no. 3, p. 507.
89. Boeve, B.F., Melatonin for treatment of REM sleep behavior disorder: response in 8 patients, *Sleep*, 2001, vol. 24, p. A35.
90. Mendelson, W.B., A critical evaluation of the hypnotic efficacy of melatonin, *Sleep*, 1997, vol. 20, p. 916.
91. Cardinali, D.P., Srinivasan, V., Brzezinski, A., and Brown, G.M., Melatonin and its analogs in insomnia and depression, *J. Pineal Res.*, 2012, vol. 52, no. 4, p. 365.
92. Guardiola-Lemaître, B., Toxicology of melatonin, *J. Biol. Rhythms*, 1997, vol. 12, no. 6, p. 697.
93. Willis, G.L., The role of ML-23 and other melatonin analogues in the treatment and management of Parkinson's disease, *Drug News Perspect.*, 2005, vol. 18, no. 7, p. 437.
94. Willis, G.L., Moore, C., and Armstrong, S.M., A historical justification for and retrospective analysis of the systematic application of light therapy in Parkinson's disease, *Rev. Neurosci.*, 2012, vol. 23, no. 2, p. 199.
95. Catalá, M.D., Cañete-Nicolás, C., Iradi, A., et al., Melatonin levels in Parkinson's disease: drug therapy versus electrical stimulation of the internal globus pallidus, *Exp. Gerontol.*, 1997, vol. 32, nos. 4–5, p. 553.
96. Willis, G.L. and Armstrong, S.M., A therapeutic role for melatonin antagonism in experimental models of Parkinson's disease, *Physiol. Behav.*, 1999, vol. 66, no. 5, p. 785.
97. Tapias, V., Cannon, J.R., Greenamyre, J.T., et al., Melatonin treatment potentiates neurodegeneration in a rat rotenone Parkinson's disease model, *J. Neurosci. Res.*, 2010, vol. 88, no. 2, p. 420.
98. Chen, H., Schernhammer, E., Schwarzschild, M.A., and Ascherio, A., A prospective study of night shift work, sleep duration, and risk of Parkinson's disease, *Am. J. Epidemiol.*, 2006, vol. 163, no. 8, p. 726.
99. Schernhammer, E.S., Rosner, B., Willett, W.C., et al., Epidemiology of urinary melatonin in women and its relation to other hormones and night work, *Cancer Epidemiol., Biomarkers Prev.*, 2004, vol. 13, no. 6, p. 936.
100. Shirani, A. and St Louis, E.K., Illuminating rationale and uses for light therapy, *J. Clin. Sleep Med.*, 2009, vol. 5, p. 155.
101. Artemenko, A.R. and Levin, Ya.I., The phototherapy of parkinsonism patients, *Zh. Nevrol. Psikhiatr.*, 1996, vol. 96, no. 3, p. 63.
102. Paus, S., Schmitz-Hubsch, T., Wullner, U., et al., Bright light therapy in Parkinson's disease: a pilot study, *Mov. Disord.*, 2007, vol. 22, no. 10, p. 1495.
103. Willis, G.L. and Turner, E.J., Primary and secondary features of Parkinson's disease improve with strategic exposure to bright light: a case series study, *Chronobiol. Int.*, 2007, vol. 24, no. 3, p. 521.
104. Witkovsky, P., Dopamine and retinal function, *Doc. Ophthalmol.*, 2004, vol. 108, no. 1, p. 17.
105. Nir, I., Haque, R., and Iuvone, P.M., Diurnal metabolism of dopamine in the mouse retina, *Brain Res.*, 2000, vol. 870, nos. 1–2, p. 118.

106. Andrade, L.A., Lima, J.G., Tufik, S., et al., REM sleep deprivation in an experimental model of Parkinson's disease, *Arq. Neuro-Psiquiatr.*, 1987, vol. 45, no. 3, p. 217.
107. Demet, E.M., Chicz-Demet, A., Fallon, J.H., and Sokolski, K.N., Sleep deprivation therapy in depressive illness and Parkinson's disease, *Prog. Neuro-Psychopharmacol. Biol. Psychiatry*, 1999, vol. 23, no. 5, p. 753.
108. Reist, C., Sokolski, K.N., Chen, C.C., et al., The effect of sleep deprivation on motor impairment and retinal adaptation in Parkinson's disease, *Prog. Neuro-Psychopharmacol. Biol. Psychiatry*, 1995, vol. 19, no. 3, p. 445.
109. Willis, G.L., Parkinson's disease as a neuroendocrine disorder of circadian function: dopamine-melatonin imbalance and the visual system in the genesis and progression of the degenerative process, *Rev. Neurosci.*, 2008, vol. 19, nos. 4–5, p. 245.
110. McMahon, D.G., Iuvone, P.M., and Tosini, G., Circadian organization of the mammalian retina: from gene regulation to physiology and diseases, *Prog. Retinal Eye Res.*, 2014, vol. 39, p. 58.
111. Tosini, G. and Menaker, M., Circadian rhythms in cultured mammalian retina, *Science*, 1996, vol. 272, no. 5260, p. 419.
112. Vanecek, J., Cellular mechanisms of melatonin action, *Physiol. Rev.*, 1998, vol. 78, no. 3, p. 687.
113. Rodnitzky, R.L., Visual dysfunction in Parkinson's disease, *Clin. Neurosci.*, 1998, vol. 5, no. 2, p. 102.
114. Sartucci, F., Orlandi, G., Lucetti, C., et al., Changes in pattern electroretinograms to equiluminant red-green and blue-yellow gratings in patients with early Parkinson's disease, *J. Clin. Neurophysiol.*, 2003, vol. 20, no. 5, p. 375.
115. Altıntaş, O., Işeri, P., Ozkan, B., and Çağlar, Y., Correlation between retinal morphological and functional findings and clinical severity in Parkinson's disease, *Doc. Ophthalmol.*, 2008, vol. 116, no. 2, p. 137.
116. Zisapel, N., Melatonin-dopamine interactions: from basic neurochemistry to a clinical setting, *Cell. Mol. Neurobiol.*, 2001, vol. 21, no. 6, p. 605.
117. Rutten, S., Vriend, C., Heuvel, O.A., et al., Bright light therapy in Parkinson's disease: an overview of the background and evidence, *Parkinson's Dis.*, 2012, article ID 767105.
118. Srinivasan, V., Cardinali, D.P., Srinivasan, U.S., et al., Therapeutic potential of melatonin and its analogs in Parkinson's disease: focus on sleep and neuroprotection, *Ther. Adv. Neurol. Disord.*, 2011, vol. 4, no. 5, p. 297.

Translated by M. Batrukova